



A Novel Stability Indicating Rp-Uplc-Dad Method for Determination of Metformin and Empagliflozin in Bulk and Tablet Dosage form

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ABSTRACT

The objective of the present work was to develop and validate a novel stability indicating RP-UPLC-DAD method for the simultaneous analysis of Metformin and Empagliflozin in bulk and tablet dosage form. This new RP-UPLC method has superior in technology to formal Reverse phase-HPLC with retention time, solvent utilization, resolution and less cost. The peak area separation was accomplished on a Waters model UPLC system equipped with PDA detector and autosampler. A volume of 5 μ L of sample and standard were injected into the column and all analytes were separated by using the mobile phase contains mixture 0.1% ortho phosphoric acid buffer (the pH was adjusted to 3.4 with 0.1 N NaOH) and methanol in the ratio 40:60% v/v at a flow rate of 0.25ml/min through C18 BEH(Ethylene Bridged Hybrid) UPLC (100mm x 2.1mm, 1.8 μ m) at 35°C column temperature and the detector wavelength was set at 254 nm. The system suitable parameters such as tailing factor, resolution and plate count of two drugs were 1.16, 1.37; 3.47; 2314.34 and 4723 respectively. Retention time and peak area of Metformin and Empagliflozin were found to be 0.882 & 3.471, 4887835 & 163463 respectively. Regression equation shows an r value (correlation coefficient) of greater than 0.999 for Metformin and Empagliflozin. Percent recovery of Metformin and Empagliflozin was found to be 99.92%-100.12% & 100.12%-100.56% respectively. Both Metformin and Empagliflozin were subjected to stress conditions such as acidic, basic, oxidative, thermal and photo degradation but substantial degradation was observed in acid studies. The newly developed RP-UPLC-DAD chromatographic method was validated with regard of system suitability, linearity, robustness, accuracy, LOD, precision and LOQ.

Keywords: Empagliflozin, Metformin, RP-UPLC, Stress studies, Validation.

INTRODUCTION

Metformin is now established as a first choice drug for all type 2 diabetes mellitus patients,

except when not tolerated or contraindicated it suppress hepatic gluconeogenesis and glucose output from liver. This is the major action responsible for lowering of blood glucose in diabetics. Enhance

insulin mediated glucose uptake and disposal in skeletal muscle and fat. Insulin resistance exhibited by type II diabetics is thus overcome¹⁻². Worldwide most of the patients with type II diabetes mellitus are commonly accomplished with a singular-agent therapy, generally metformin and it is a good and most commonly used antihyperglycemic agent which enhance the glucose margin in type 2 diabetes patients, lowering both postprandial and basal plasma glucose. Metformin molecular formula $C_4H_{11}N_5 \cdot HCl$ was and molecular weight 165.62³⁻⁶ it is a white crystalline compound, Structure of Metformin shown in Fig 1.

Practically all the glucose particles filtered at the glomerulus and it is reabsorbed in the proximal tubules. The major transporter which accomplishes this is sodium glucose cotransporter-2, whose inhibition induces glycosuria and lowers blood glucose in type 2 DM, as well as causes weight loss. This SGLT-2 inhibitor Empagliflozin has been recently tested in type 2 DM patients. After single daily dose it lowers blood glucose levels⁷⁻⁹.

Empagliflozin chemically, n-Glucitol, 1, 5-anhydro-1-C-[4-chloro-3-[[4-[[[(3S)-tetrahydro-3-furanyl]oxy]phenyl]methyl]phenyl]-, (1S). SGLT2 was highly stated in kidney. Empagliflozin plays an important role in good transporter and the reabsorption of glucose from the glomerular filtrate sent back into the circulation¹⁰⁻¹¹. Structure of Empagliflozin was shown in Fig 2.

Fast estimation of Metformin and Empagliflozin in with more difference in label claims (Empagliflozin for 5 mg and Metformin for 500

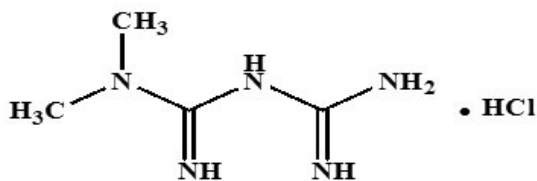


Fig. 1: Structure of Metformin

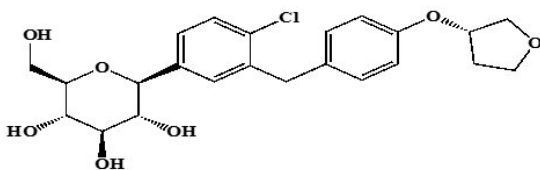


Fig. 2: Structure of Empagliflozin

mg) with short run time is a good challenge. For UPLC based analysis, the method of decreasing assays time effectively while resolving analytes from degraded products is often attained with column with small particles.

Literature survey brings out that Empagliflozin is determined by UV spectrophotometric method¹²⁻¹⁴, RP-HPLC method¹⁵⁻¹⁸, very less chromatographic methods were available for simultaneous determination of Metformin and Empagliflozin in bulk and tablet dosage form by using RP-HPLC¹⁹⁻²⁴, UPLC²⁵.

The analytical method development and validation of Metformin and Empagliflozin using Gas liquid /Mass spectrometry and Liquid chromatography /Mass spectrometry were expensive and very delicate methods as compared with the UPLC for general analysis, hence it needs to use this UPLC –DAD technique for determination of stability studies by using RP-UPLC-DAD method Metformin and Empagliflozin in bulk and tablet dosage form.

EXPERIMENTAL

Chemicals and reagents

Qualified Empagliflozin standard was kindly obtained by Manus akkteva (Ahmadabad, Gujarat, India). Metformin was kindly obtained by Aurobindo pharmaceuticals Ltd. (Hyderabad, Telengana, India). Synjardy® (Empagliflozin -5mg and Metformin-500 mg) (USA) were purchased from local market. Orthophosphoric acid AR grade was purchased from Merck (Darmstadt, Germany). HPLC grade solvent methanol was obtained from JVR fine chemicals ltd (Hyderabad, Telengana, India). Millipore -Q® type 1 ultrapure water system (Sartorius, Germany), vacuum pump was purchased from PCI Analytics Pvt, Ltd (Mumbai, India) were also used.

Instrumentation and Equipments

The UPLC waters system was used for the stability indicating method development and method validation. It consists of binary solvent manager with connected with photo diode array detector (MA, USA) controlled with Empower software and auto sampler and auto injector. C18 BEH (Ethylene Bridged Hybrid) UPLC (100mm x 2.1mm, 1.7µm) column was used

from waters. Photo stability studies were performed in photo stability chamber (Osworld scientific equipment pvt.Ltd, Mumbai, India). Thermal stability studies were carried out in a dry air oven (Newtronic life care, Mumbai, India).

Solution preparations

Preparation of mobile phase

0.1% ortho phosphoric acid buffer solution was accurately prepared by taking 1 ml ortho phosphoric acid in 1000 ml volumetric flask and

dissolved it in water then made up to the mark by adjust the pH of solution to pH =3.4 with 0.1 N NaOH solution. Then filtered the resulting solution through 0.45 μ filter under vacuum filtration. Mixture of ortho phosphoric acid buffer and methanol in the ratio 40:60% v/v was taken in a flask, degassed in ultrasonic water bath for 5 minutes allowed to cool at room temperature and then filtered through 0.45 μ filter under vacuum filtration. This prepared mobile phase was used as diluent.

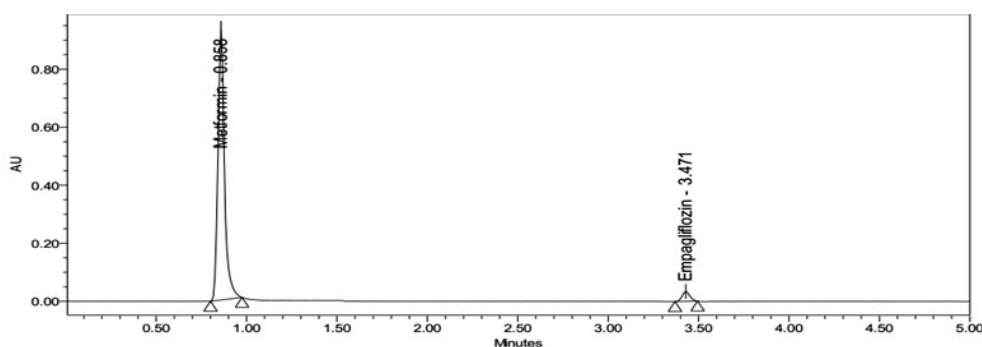


Fig. 3: Optimized standard chromatograms of Metformin and Empagliflozin

Table 1: System suitability results of Metformin and Empagliflozin

S. no	Retention time	Peak	USP plate area	USP tailing count	Resolution
Metformin	0.882	4887835	2314.34	1.16	3.471
	0.888	4877016	2213.15	1.19	3.472
	0.885	4854854	2330.21	1.18	3.475
	0.889	4870934	2332.58	1.15	3.476
	0.895	4866954	2256.65	1.17	3.474
	0.889	4850952	2254.32	1.16	3.472
Mean		4868090			
S.D		13768.2			
% RSD		0.28			
Empagliflozin	3.471	163463	4723.0	1.36	
	3.472	162404	4277.64	1.37	
	3.475	156082	4463.81	1.36	
	3.476	162766	4655.72	1.37	
	3.474	166211	4682.24	1.36	
	3.472	159403	4407.81	1.36	
Mean		161721.5			
S.D		3519.3			
% RSD		0.7			

Precision

Preparation of standard solution

Weigh accurately and transfer 100 mg Metformin and 10 mg Empagliflozin working standard into a 10 ml clean dry volumetric flask and add about 7 ml of and sonicated to dissolve it totally and make up the volume to the up mark with the diluent. (Stock solution). After that pipette out 0.3 ml of the above solutions into a 10ml dry volumetric flask and dilute up to the mark with diluent.

Preparation of sample solution

Weigh accurately 10 tablets crush in mortar and pestle and transfer equivalent to 500mg

Metformin and 5mg Empagliflozin sample into a 10 ml clean dry volumetric flask add about 7 ml of diluent and sonicated it up to 30 mins to dissolve it completely and make up the volume up to the mark with with the diluent. Then it is filtered through 0.44 micron injection filter. After that pipette out 0.3 ml of Metformin and Empagliflozin from the above stock solution into a 10ml dry volumetric flask and dilute up to the mark with diluent and inject 5 mL of the standard and sample into the RP-UPLC system.

Chromatographic conditions

Chromatographic column used was C18

Table 2: Precision study of Metformin and Empagliflozin

Name of drug	Amount applied ($\mu\text{g/ml}$)	Repeatability (Mean peak area \pm S.D)	% RSD	Intermediate precision (Mean peak area \pm S.D)	% RSD
Metformin	15	2304436.5 \pm 41123.20	1.78	2304526.5 \pm 41137.30	1.78
	30	2304436.5 \pm 39708.99	1.72	2304576.5 \pm 41208.06	1.78
	45	2309936.5 \pm 46072.95	1.99	2305076.5 \pm 40500.90	1.75
Empagliflozin	25	92605.0 \pm 711.3	0.76	92563.5 \pm 639.3	0.69
	50	92605.5 \pm 713.4	1.16	92613.5 \pm 569.2	0.61
	75	92613.5 \pm 710.6	1.18	92663.5 \pm 639.9	0.69

Table 3: Percentage recovery of Met formin and Empagliflozin

Name of drug	Spiked level (%)	Amount added	Amount recovered	% Recovery	% RSD
Metformin	80	48.51	49.08	100.12	0.42
	100	98.5	99.02	100.06	0.34
	120	147.1	148.32	99.92	0.76
Empagliflozin	50	4.62	4.95	100.14	0.39
	100	9.82	9.94	100.56	0.68
	150	14.24	14.56	100.36	0.82

Table 4: Linearity data of Metformin and Empagliflozin

S.no	Metformin Conc. ($\mu\text{g/ml}$)	Peak area	Empagliflozin Conc. ($\mu\text{g/ml}$)	Peak area
1	25	1737618	15	99860
2	50	3100784	30	166350
3	75	4447974	45	230484
4	100	5851768	60	289354
5	125	7002620	75	351112
Slope	53124		4170.1	
Y-intercept	443856		39780	
Correlation coefficient	0.999		0.999	

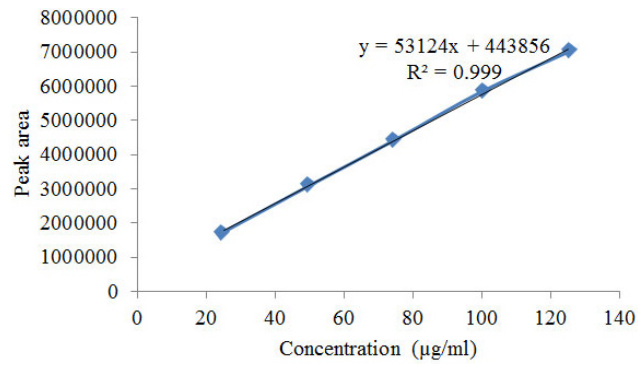


Fig. 4: Linearity curve of Metformin

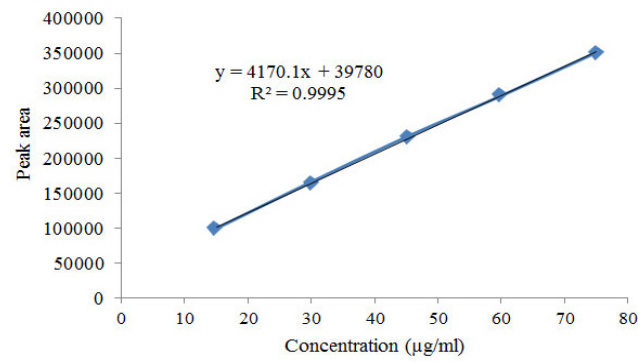


Fig. 5: Linearity curve of Empagliflozin

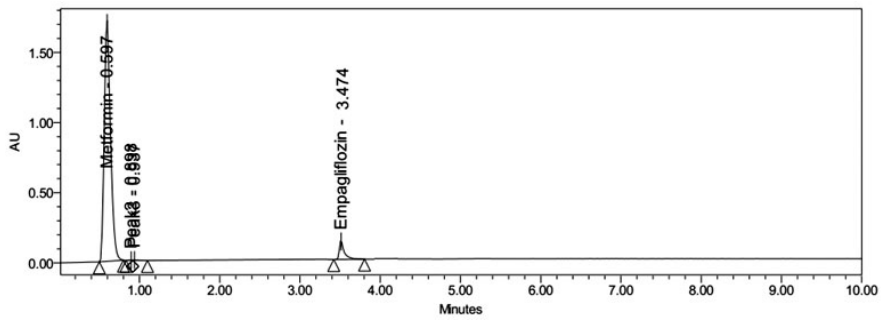


Fig. 6: Acid degradation chromatogram of Metformin and Empagliflozin

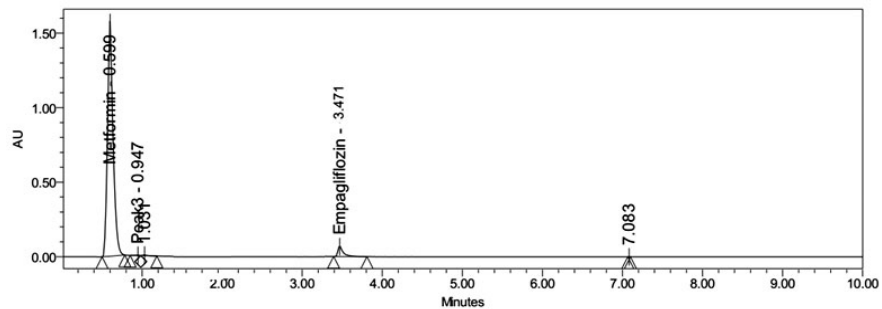


Fig. 7: Base degradation chromatogram of Metformin and Empagliflozin

BEH UPLC, 100mm x 2.1mm with particles 1.8 μ m. The mobile phase consist of mixture of 0.1% ortho phosphoric acid buffer and methanol in the ratio 40:60% v/v with a flow rate of 0.25ml/min, monitored at a wavelength of 254 nm and it maintained at 35 °C with a injection volume of 5 μ l.

Method validation

System suitability parameters

A system suitability test method was expressed based on the parameters results obtained in different chromatograms. The concentration of standard solution was 300 μ g/ml for Metformin

Table 5: Robustness of Metformin and Empagliflozin

S.no	Metformin peak area	Metformin %RSD	Metformin USPplate count	Empagliflozin Peak area	Empagliflozin % RSD	Empagliflozin USPplate count	
Flow rate	0.80	486988.5 \pm 631.4	0.12	2213.24	226543 \pm 2626	1.15	3711
	1.00	486938.5 \pm 702.1	0.14	2219.12	227543 \pm 1211	0.53	3725
	1.20	487888.5 \pm 782.7	0.16	2268.32	226635 \pm 2919	1.28	3752
Mobile phase composition							
	30:70	487943.5 \pm 704.9	0.14	2316.2	227600 \pm 1979	0.86	3893
	40:60	488048.5 \pm 853.4	0.17	2321.0	227100 \pm 1272	0.56	3819
	50:50	487888.5 \pm 782.7	0.16	2372.2	226100 \pm 2687	1.18	3833
Column temperature							
	30°C	478048.5 \pm 853	0.17	2318.3	226600 \pm 1979	0.87	3963
	35°C	477548.5 \pm 1560	0.32	2324.1	227635 \pm 1505	0.66	3890
	45°C	475548.5 \pm 1267	0.26	2352.3	226377 \pm 1665	0.73	3933

Table 6: Degradation results of Metformin and Empagliflozin

S.no	Metformin area	Metformin % degraded	Metformin % recovery	Empagliflozin area	Empagliflozin % degraded	Empagliflozin % recovery
Acid degradation	8490245	90.45	9.55	659212	90.78	9.22
Base degradation	6413362	91.02	8.98	413095	92.12	7.88
Peroxide degradation	29871421	96.45	3.55	453286	95.21	4.79
Thermal degradation	8123070	96.53	3.47	328834	96.18	3.82
Photo degradation	8232042	96.82	3.18	432872	96.45	3.55

and 30 µg/ml for Empagliflozin respectively. The efficiency of column resolute from the analyte peak >20, 000, the tailing factor <5.0% and resolution between peaks of two drugs should be >1.5.

Precision

Precision was evaluated by studying the intermediate precision and repeatability. Precision of each sample was determined by six replicate injections at a concentration of 100µg/ml of Metformin and 10 µg/ml of Empagliflozin respectively.

Accuracy

To evaluate the accuracy of proposed method, recovery studies were conducted, it is determined in three times at three different levels of concentration i.e. 80%, 100%, 120% in tablet dosage and bulk form.

Linearity

The linearity of analytical method was determined by measuring different concentrations of standard solutions to Metformin (25-125 µg/ml) and Empagliflozin (15-75 µg/ml). The calibration curve was obtained by plotting concentration of sample solutions on X-axis and mean peak areas on Y-axis.

Sensitivity

Sensitivity of the proposed analytical method was determined in terms of limit of limit of quantitation (LOQ) and detection (LOD). $LOQ = 10 \times ASD/S$ and $LOD = 3.3 \times ASD/S$. In order to determine both parameters, concentration in the lower part of the linear range of calibration curve were used.

Robustness

The robustness method measures the method capability to it will not affected by very small, but if any deliberate changes in optimized chromatographic conditions was examined by testing influence of small deliberate changes in, change in mobile phase composition (90% to 110%), flow rate (± 0.2 ml/min), column temperature ($\pm 5^\circ\text{C}$) using 100µg/ml Metformin and 10 µg/ml Empagliflozin.

Stability studies

In this stability studies, Metformin and Empagliflozin were exposed to different chemical and physical degradation conditions such as 0.1 N HCl (acid degradation), 0.1% N NaOH (base degradation), 30% H_2O_2 (oxidative degradation), heat (thermal degradation) and Ultra violet light (radiation decay) for defined time intervals, and then diluted similar as standard stock dilution, and then representative chromatograms were obtained. The percentage of degradation compounds were calculated from the peak areas of the Metformin and Empagliflozin chromatograms. In the study of base or acid degradation, an amount of equivalent to powdered sample to 100mg Metformin and 10 mg Empagliflozin was transferred into 10 ml dry volumetric flask and add 3 ml of freshly prepared 0.1 N NaOH or 0.1 N HCl. then shaken well and allowed for 24 hours at a temperature of 60°C. Then filtered the total solution through 0.45µ filter into 10 ml standard flasks and neutralized the un reacted acid or base i.e. 0.1 N HCl or 0.1N NaOH and made up to the mark. In case of oxidative degradation same amount of sample was transferred into 10 ml volumetric flask, add 3 ml of freshly prepared 30%

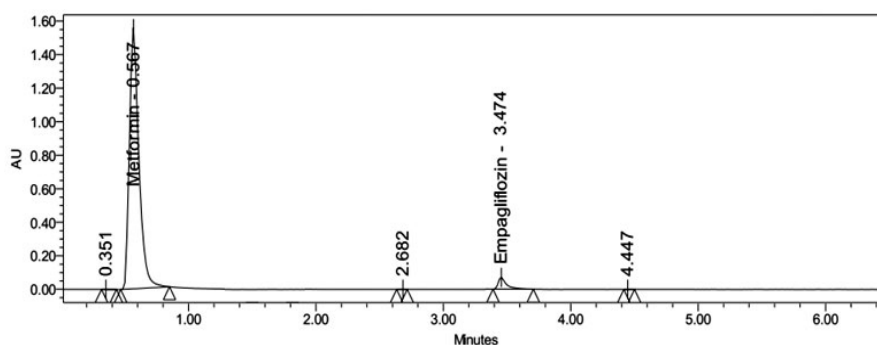


Fig. 8: Oxidative degradation chromatogram of Metformin and Empagliflozin

H₂O₂ and kept at 75°C for 48 hours and filtered the total solution through 0.45µ filter and made up to the mark. In the study of thermal decay or Ultra violet light -light degradation, precisely same quantity of sample was transferred into clean and dry watch glass, placed in a hot air oven at 110 °C. In the study of photo degradation precisely same amount of sample was transferred into 10 ml volumetric flask kept at sunlight for 48 hours and then made up to the mark with diluents.

RESULTS AND DISCUSSION

UPLC method development

UPLC is a new analytical technique used for determination of pharmaceutical formulations and separations mostly in combined drugs and dosage forms. The main plan of this work was to develop a novel stability indicating RP-UPLC method for estimation of Metformin and Empagliflozin. Initial trials were conducted during UPLC method development for optimizing different parameters on the system suitability of the method.

After trying different columns like, cyano, PFP, C8 the last chance of the stationary phase that gave a good resolution and eluting all peaks with better separation was reverse phase C18 BEH UPLC (100mm x 2.1mm 1.8µm particle size) column, this column was only showing retention time less than 1.5 minutes. The organic solvents acetonitrile and methanol were tried for analytical method development. Compare with acetonitrile, methanol was showing good peak separations and less column back pressure. As we need to analysis of compounds, tried with different mobile phases like phosphate buffer, formic acid, trifluoroacetic acid ortho phosphoric acid. In acid hydrolysis drugs were

showing good peak separation and shape. Finally the good results were obtained by use of mixture of 0.1% ortho phosphoric acid (pH of the buffer adjusted to 3.4 with 0.1 N NaOH) and methanol in the ratio 40:60% v/v at a flow rate of 0.25ml/min through C18 BEH (Ethylene Bridged Hybrid) UPLC (100mm x 2.1mm, 1.8µm) at 35 °C column temperature and the detector wavelength was set at 254 nm. Under the optimum chromatographic conditions, the retention times obtained for Metformin and Empagliflozin were 0.882 and 3.471 min, respectively. Optimized standard chromatogram for Metformin and Empagliflozin were expressed in figure 3.

Method validation

System suitability

System suitability parameters such as resolution, tailing factor, plate count and resolution for two drugs were found to be 3.47; 1.16 and 1.37; 2314.34 and 4723 respectively. Peak area and retention time of Metformin and Empagliflozin were found to be 4887835 & 163463, 0.882 & 3.471 respectively. The results of system suitability were expressed in Table 1.

For repeatability and intermediate precision assessment, %RSD was calculated. All the samples exhibited RSD values < 2 % confirming that the analytical method was precise. The results of precision were expressed in Table 2.

Accuracy

The acceptable percentage recovery of Metformin and Empagliflozin in bulk and tablet dosage form ranged from 99.92%-100.12 % and 100.14%-100.56 % respectively. The results are tabulated in Table 3.

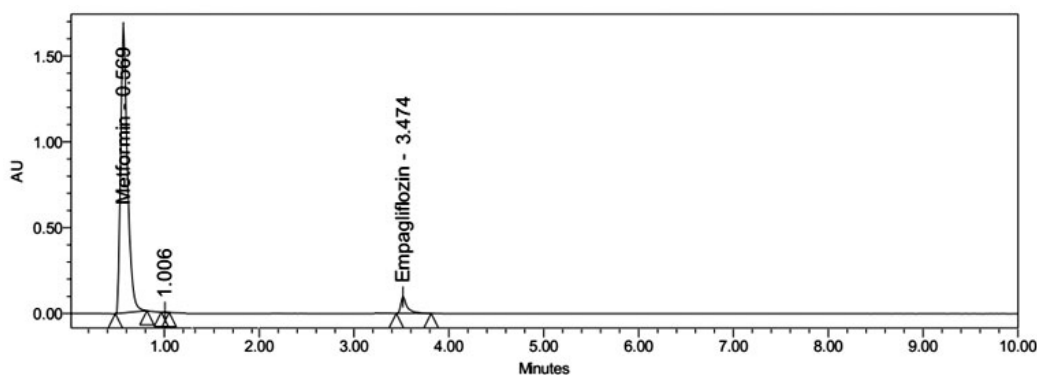


Fig. 9: Thermal degradation chromatogram of Metformin and Empagliflozin

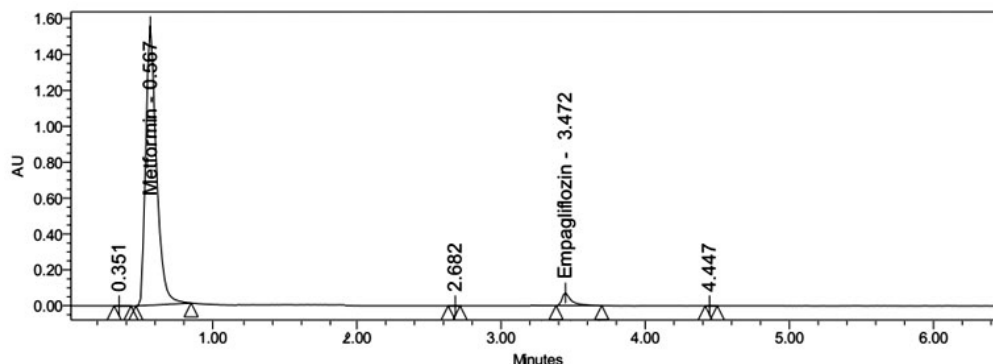


Fig.10: Photo degradation chromatogram of Metformin and Empagliflozin

Linearity

The linear regression equation and correlation (r) for Metformin and Empagliflozin were $y = 53124x + 443856$, 0.999 and $y = 4170.1x + 39780$, 0.999 respectively. Linearity curve of both drugs were expressed in Table 4, Fig 4 and 5.

Sensitivity

The limit of detection and limit of quantification of Metformin and Empagliflozin were found to be 0.072 $\mu\text{g/ml}$, 0.330 $\mu\text{g/ml}$ and 0.016 $\mu\text{g/ml}$, 0.964 $\mu\text{g/ml}$, respectively.

Robustness

Robustness of the method was expressed by % RSD values of Metformin and Empagliflozin peak areas as quantitative responses. All values were found to be less than 2%, indicates that method was robust. Results are shown in table 5.

Stability studies

Degraded products were not found in oxidative, photolytic, thermal condition which confirms that Metformin and Empagliflozin were stable to oxidative, photolytic, thermal conditions. The drugs were found to be liable to acid, base hydrolysis as a total of 9.55, 9.22 & 8.98, 7.88 degradation was found respectively. Degradation

details were expressed in table 6. Acid, base, oxidative, thermal, photolytic chromatograms were shown in figures 6-10.

CONCLUSION

The proposed novel stability indicating isocratic RP-UPLC-DAD method was developed and validated for determination of Metformin and Empagliflozin in bulk and tablet dosage. Validation proves that the proposed method is good and satisfactory for determination of Metformin and Empagliflozin in tablet dosage and bulk form for accuracy, specificity, LOQ, LOD, precision, linearity and robustness. The proposed method is reliable, simple, economical and adequate for use in routine drug quality control analysis in laboratories and pharmaceutical industry. Stability studies were done to assess the stability of compound and prove the stability indicating nature of developed RP-UPLC method.

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