



Prenylated Xanthone and Rubraxanthone with Antiplatelet Aggregation Activity in Human Whole Blood Isolated from *Garcinia griffithii*

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ABSTRACT

Phytochemical studies on the methanolic extract of the leaves of *Garcinia griffithii* (Guttiferae) afforded two known xanthones derivatives, 1,3,5,6-Tetrahydroxy-7-(3-methylbut-2-enyl)xanthone (1) and Rubraxanthone (2). The structures of these xanthones derivatives were determined on the basis of extensive spectroscopic data interpretation and comparison with data reported in the literature. In an in vitro test, compound 1 and 2 showed the ability to inhibit platelet aggregation in human whole blood which induced by arachidonic acid (AA), adenosine diphosphate (ADP) and collagen.

Key words: *Garcinia griffithii*, Platelet aggregation, Impedance Method,
NMR spectroscopy, HMBC correlations.

INTRODUCTION

Plant secondary metabolites are a group of naturally occurring compounds classes biosynthesized by differing biochemical pathways whose plant content and regulation is strongly susceptible to environmental influences and to potential herbal predators. From ancient to modern times, herbs and plants have been used as medicinal agents, first only on a folkloric basis and later developed on a scientific basis. Natural products of plant origin offered a wide variety of bioactive compounds that could meet the demand for base

compounds of drugs¹. A large proportion of the drugs used in modern medicine were either directly isolated from plants or synthetically modified from a lead compound of natural origin. Drug discovery is an iterative process of lead discovery (i.e. isolation of bioactive lead compounds from these natural sources) coupled with lead improvement (rational design and synthesis of new analogues to improve pharmacological profiles) and natural products play an important role in drug development programs of pharmaceutical industry. Over 50% of all modern clinical drugs are of natural product origin and higher plants as source of medicinal compounds continue

to play a dominant role in maintenance of human health since antiquities²⁻⁶.

Guttiferae (Clusiaceae) plant species have been proven a rich source of bioactive compounds. The family *Guttiferae* numbers over 1000 species mainly confined to the tropics. *Garcinia* belongs to *Guttiferae* (Clusiaceae) family and is a large genus of polygamous trees or shrubs, distributed in the tropical Asia, Africa and Polynesia 9-11. The genus *Garcinia* consists of 180 species and about 30 species occur in India. For the last several years, chemical investigations of various *Garcinia* species attracted the attention of chemists and have been extensively investigated from phytochemical and biological points of view due to the interesting biological properties of many of its species. *Garcinia* species are a rich source of secondary metabolites including xanthenes, flavonoids, benzophenones, lactones and phenolic acids and a variety of simple and complex bioactive molecules have been isolated from the different parts of African and south east Asian *Garcinia* species. Phytochemical investigation of the fruits, stem barks, seeds, leaves and roots of different *Garcinia* species have resulted in the isolation of complex molecules including xanthenes, prenylated xanthenes, prenylated benzophenones, flavonoids, lactones and triterpenes 9-19. As part of a phytochemical study of South East Asian plants, herein we report the purification, structure characterization, and biological testing of two known xanthenes derivatives 1 and 2 (Figure 1), which are common to *G. griffithii* collected from Negeri Sembilan, Malaysia.

MATERIAL AND METHODS

General experimental procedures

Melting points were determined on a Yanaco MP-S3 apparatus. UV spectra were measured on a Shimadzu UV 240 spectrophotometer. JASCO DIP-360 Digital polarimeter was used to measure the optical rotations in chloroform by using 10 cm cell tube. FTIR-8900 Spectrophotometer was used to record IR spectra in CHCl_3 . The ¹H-NMR and ²D NMR spectra were recorded on a Bruker Avance III 500 Ascend spectrometer using BBO probe, while ¹³C-NMR spectra were recorded on Bruker Avance III 500 Ascend spectrometer operating at 125 MHz using CDCl_3 as solvent. Chemical shifts

were reported in δ (ppm), relative to SiMe₄ as internal standard, and coupling constants (J) were measured in Hz. The EI-MS and HREIMS were measured on Jeol HX 110 mass spectrometer. TLC was performed on Si gel precoated plates (PF254, 20 × 20, 0.25 mm, Merck, Germany). Ceric sulphate in 10% H₂SO₄ spraying reagent was used for the staining of compounds on TLC. All reagents used were of analytical grades 18.

Plant material

The leaves of *Garcinia griffithii* (*Guttiferae*) was collected from Pasoh, Negeri Sembilan, Malaysia in September 2012. A voucher specimens (MT29 & MT31) were deposited in the herbarium of the Herbarium of the Forest Research Institute Malaysia (FRIM), Kepong, Malaysia.

Extraction

Dried leaves of *Garcinia griffithii* (*Guttiferae*) was extracted under reflux with MeOH for 6-8 hrs and filter it. Obtained MeOH extract was fractionate through vacuum liquid chromatography (VLC) technique using different polarity. Evaporation of the respective solvents and purified the compounds through Preparative thin layer chromatography 18.

Impedance Method

Whole blood (1ml) diluted with buffer saline (1:1) was incubated with samples 5 μ l, in DMSO at 37° C for 2 min. After which collagen (2 μ g/ml), ADP (10 μ M), AA(0.5 mM) was added to initiate aggregation. The platelet aggregation was measured by whole blood Lumi-Aggregometer (Chrono-Log Corp. Haver-town, PA). The results of aggregation in whole blood was determined after 5 min. as the increase in impedance across a pair of electrodes placed in the blood sample by comparison to that of control group impedance. To eliminate the effect of the solvent on the aggregation, blood with 0.5% DMSO was used as the control¹⁸. The percentage inhibition of platelet aggregation was calculated as:

$$\% \text{ Inhibition} = 1 - \frac{(\text{aggregation of sample})}{(\text{aggregation of control})} \times 100$$

RESULTS AND DISCUSSION

Phytochemical investigation

Dried leaves of *Garcinia griffithii* (*Guttiferae*)

Table 1: Percentage inhibition of isolated compounds (1-2) from Guttiferae species on platelet aggregation in human whole blood induced by arachidonic acid (AA) (0.5mM), collagen (2µg/mL), and adenosine diphosphate (ADP) (10µM)

Compounds name	Concentration (µg/ml)	AA	COLLAGEN	ADP
1,3,5,6-Tetrahydroxy-7-(3-methylbut-2-enyl)xanthon (1)	100	100±0.00	62.3±0.71c	100±00a
	50	62.5 ±0.7	42.5± 1.4	65.6±0.45
	25	45.1± 1.4	22.8± 0.7	30.8±0.71
	12.5	25.2± 0.7	6.7± 1.3	16.1± 0.7
Rubraxanthone (2)	100	57.8±0.7	100±00a	100±00a
	50	44.6±0.48	91.3±1.5	
	66.6±0.45			
	25	24.1±1.2	65.3±061	30.4±0.85
	12.5	13.2±0.58		
	29.2±0.48			

Table 2: IC50 values (µM) of isolated compounds (1-2) on platelet aggregation induced by arachidonic acid (AA) (0.5mM), Collagen (2µg/ml), and adenosine diphosphate. (ADP) (10µM)

Compounds	AA	Collagen	ADP
1,3,5,6-Tetrahydroxy-7-(3-methylbut-2-enyl)xanthon(1)	211.1±2.9	48.1±2.7	84.6±2.1
Rubraxanthone (2)	114.9±3.1	229.2±5.1	107.4±4.8
Aspirin	27.5±2.8	-	-

was extracted under reflux with MeOH for 6-8 hrs and filter it. Obtained MeOH extract was fractionate through vacuum liquid chromatography (VLC) technique using different polarity. Phytochemical investigations have revealed two known xanthenes derivatives, 1,3,5,6-Tetrahydroxy-7-(3-methylbut-2-enyl)xanthon (1) and Rubraxanthone (2) (Figure 1). All spectroscopic data described below were showed excellent agreement with the previously published results²⁰⁻²². These structures were confirmed by further analysis of the HMBC correlations as shown in Figure 2 and 3.

1,3,5,6-Tetrahydroxy-7-(3-methylbut-2-enyl)xanthon (1) was obtained as a yellow faint powder (11 mg); EIMS for C₁₈H₁₆O₆ m/z (rel.int.): 328[M]⁺; ¹H NMR (CD₃OD, 500 MHz): δ 1.664, 1.786, 3.301, 3.314, 4.884, 5.226, 5.240, 5.255, 6.473, 6.854, 6.872, 7.563, 7.581; ¹³C NMR (CD₃OD, 150 MHz): δ 16.733, 21.001, 24.788, 93.132, 101.847, 110.670, 112.372, 113.853, 116.350, 122.462,

130.746, 132.380, 146.428, 151.574, 155.983, 160.289, 163.248, 180.671; APT (CD₃OD, 150 MHz): 18.0346, 26.0938 (Me × 2), 94.4676, 113.7592, 117.6359, 123.6918 (CH₄), 22.2860 (CH₂), 103.1002, 111.9776, 115.0591, 132.1181, 133.723, 147.7166, 153.0461, 157.2899, 161.5412, 164.6074, 181.9876 (C_x11)²⁰⁻²².

Rubraxanthone (2) was obtained as a yellow crystal (11 mg); EIMS for C₂₄H₂₆O₆ m/z (rel.int.): 410[M]⁺; ¹H NMR (CD₃OD, 500 MHz): δ 1.51 (3H,s,3'-Me), 1.54(3H,s,8'-Me), 1.80(3H,s,3'-Me), 1.95(2H,t,4'-CH₂), 2.02(2H,m,5'-CH₂), 3.76 (3H,s, OCH₃), 4.04 (2H,d,1'-CH₂), 5.01(1H, t, 2'H), 5.21(1H, t, H_{6'}), 6.09(1H,d,2,1,H₇), 6.18(1H,d,2,1,H₅); ¹³C NMR (CD₃OD, 150 MHz): δ 181.8 (C₉), 164.7 (C₈), 163.5 (C₆), 157.2(C_{10a}), 156.8(C_{4a}), 155.5(C₃), 143.7(C₂), 137.4(C₁), 134.2(C_{3'}), 130.8(C_{7'}), 124.2(C_{6'}), 124.0(C_{2'}), 110.9(C_{9a}), 102.7(C_{8a}), 97.6(C₇), 92.8(C₅), 39.6(C_{4'}), 26.3(C_{1'}), 25.8(10'), 16.5(7'-Me), 15.4(3'-Me)²⁰⁻²².

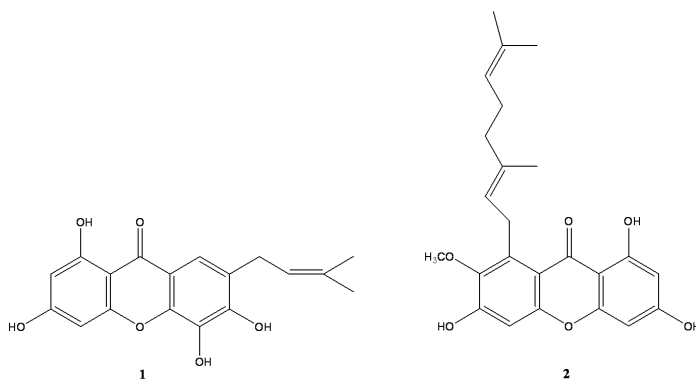


Fig. 1: Chemical structures of secondary metabolites 1 and 2 isolated from leaves of *Garcinia griffithii* (Guttiferae)

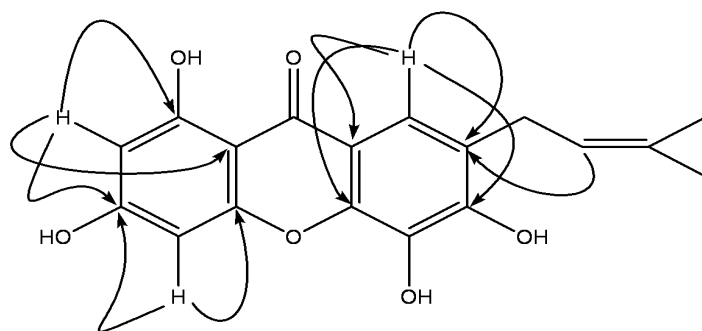


Fig. 2: HMBC correlations of compound 1

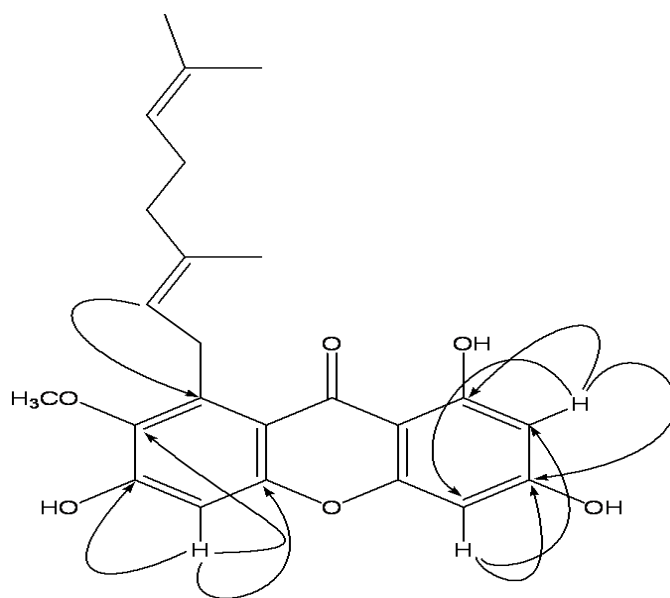


Fig. 3: HMBC correlations of compound 2

Antiplatelet aggregation activity

The effect of the isolated secondary metabolites (1-2) from *Garcinia griffithii* (Guttiferae) has been investigated against platelet aggregation test. Each sample was measured in triplicate one-way analysis of variance (ANOVA) was used for multiple comparison, the IC₅₀ values, that is, the concentration of the compounds required to inhibit aggregation by 50% ,were obtained from at least three determinations. Methanol extracts of *Garcinia griffithii* (Guttiferae) showed strong antiplatelet aggregation activity at 100µg/ml in human whole blood in vitro. Both 1, 3, 5, 6-Tetrahydroxy-7-(3-methylbut-2-enyl)xanthone (1) and Rubraxanthone

(2) showed marked inhibitory effect on platelet aggregation caused by the three inducers. The IC₅₀ value for 1,3,5,6-Tetrahydroxy-7-(3-methylbut-2-enyl)xanthone (1) was 211.1±2.9 AA, 48.1±2.7 Collagen, 84.6±2.1 ADP and the IC₅₀ value for Rubraxanthone (2) was 114.9±3.1 AA, 229.2±5.1 Collagen, 107.4±4.8 ADP, respectively (table 1 & table 2).

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REFERENCES

1. S. Kumar, S. Sharma, S. K. Chattopadhyay, *Fitoterapia*. **89**: 86-125 (2013).
2. J.T. Baker, R.P. Borris, B. Carte, G.A. Cordell, D.D. Soejarto, G.M. Cragg, *J Nat Prod*. **58**: 1325–57 (1995).
3. G.A. Cordell. *Phytochem*. **40**: 1585-612 (1995).
4. M. Stuffness, J. Douros, *J Nat Prod*. **45**: 1-14 (1982).
5. G.J. Bennett, H.H. Lee, *Phytochem*. **28**: 967-98 (1989).
6. The Wealth of India; (Raw Materials), Vol IV. New Delhi, India: CSIR; **99**: (1985).
7. V. Babu, S.M. Ali, S. Sultana, M. Ilyas. *Phytochem*. **27**: 3332–5 (1988).
8. K.R. Gustafson, J. W. Blunt, M.H.G. Munro, R.W. Fuller, T.C. Mckee, J.H. Cardellina II. *Tetrahedron*. **48**: 10093-102 (1992).
9. O. Thoison, J. Fahy, V. Dumontet, A. Chaironi, C. Riche, M.V. Tri, *J Nat Prod*. **63**: 441-6 (2000).
10. H. Minami, M. Kinoshita, Y. Fukuyama, M. Kodama, T. Yoshizawa, M. Suguira, *Phytochem*. **36**: 501-6 (1994).
11. P.G. Waterman, E.G. Crichton, *Planta Med*. **40**: 351-5 (1980).
12. P.G. Waterman, E.G. Crichton, *Phytochem*. **19**: 2723-6 (1980).
13. J. Kosin, N. Ruangrunsgi, C. Ito, H. Furukawa. *Phytochem*. **47**: 1167-8 (1998).
14. M. linuma, T. Ito, R. Miyake, H. Tosa, T. Tanaka, V. Chelladurai, *Phytochem*. **47**: 1169-70 (1998).
15. J. Asano, K. Chiba, M. Tada, T. Yoshi, *Phytochem*. **41**: 815-20 (1996).
16. V. Rukachaisirikul, M. Kamkaew, D. Sukavisit, S. Phongpaichit, P. Sawangchote, W.C. Taylor, *J Nat Prod*. **66**: 1531-5 (2003).
17. K. Likhitwitayawuid, T. Phadungcharoen, J. Krungkrai. *Planta Med*. **64**: 70-2 (1998).
18. K.A.A. Alkadi, A. Adam, M. Taha, M.H. Hasan, S.A.A. Shah, *Orient. J. Chem*. **29**(3): 871-875 2013.
19. A.E. Hay, M.C. Aumond, S. Mallet, V. Dumontet, M. Litaudon, D. Rondeau, *J Nat Prod*. **67**: 707-9 (2004).
20. C.M. Teng, C.N. Lin, F.N. Ko, K.L. Cheng, T.F. Huang, *Biochem Pharmacol*. **38**: 3791-3795 (1989).
21. C.N. Lin, S.S. Liou, F.N. Ko, C.M. Teng, *J Pharm Sci*. **82**: 11-16 (1993b).
22. M.I. Chung, J.R. Weng, J.P. Wang, C.M. Teng, C.N. Lin, *Planta Med*. **68**: 25-29 (2002).