



HPLC Method of Analysis for Determination and Standardization of Luteolin and Vanillic acid in Dry Extract of *Paronychia argentea* Lam.

HASAN Y. MUTI^{1*} and SULEIMAN OLIMAT²

¹Faculty of Pharmacy and Medical Sciences, Al-Ahliyya Amman University, Amman, Jordan.

²Faculty of Pharmacy, Jordan University of Science and Technology, Irbid, Jordan.

*Corresponding author E-mail: h.muti@ammanu.edu.jo; iris.co@orange.jo

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ABSTRACT

The main objective of this study was to establish a chromatographic method for analysis, determination and standardization of the two main components vanillic acid and luteolin as major components in *Paronychia argentea* Lam dry extract. This analytical method was designed to be a simple and fast with an appropriate separation of the two main components of the extract. High pressure liquid chromatography (HPLC) method of analysis was developed to quantitatively determine, identify and standardize the two main active constituents in the pharmaceutical dry extract against luteolin and vanillic acid as primary reference standards as it is the major active constituents of the dry extract of *P. Argentea*, where the linearity obtained was higher than $R^2 = 0.99981$ and 0.99908 respectively. Although the method was proven to be suitable, further specific analysis validation was conducted to include the following: linearity, precision, range, limit of detection, limit of quantitation and filter compatibility. The luteolin and vanillic acid were completely separated from the other components in the herbal dry extract with an R_f value of 1.3 and 5.7 min. respectively. The concentration of Luteolin is 0.4% while vanillic acid content is 0.1% in the dry extract.

Keyword: *Paronychia argentea*, HPLC, Analysis, Vanillic acid, Luteolin, Extract, Quantitation, Validation, Linearity, Precision and Repeatability.

Abbreviations: HPLC, High Pressure Liquid Chromatography, AchE, Acetyl Cholinesterase, RS, Reference Standard, LOD, Limit of Detection, LOQ, Limit of Quantitation.

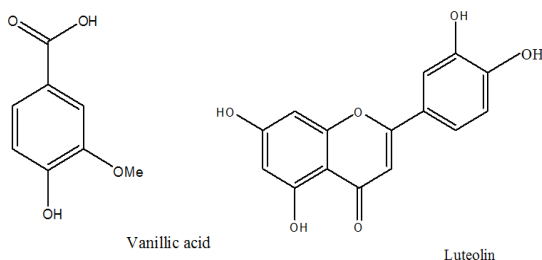
INTRODUCTION

Paronychia argentea Lam. (Locally known as Rijl El Hamameh) is a perennial herb, distributed widely throughout in Jordan¹. Several studies showed that *Paronychia argentea* has hypoglycemic activity²⁻⁴, and it has been proved to be useful as gastric analgesic, bladder, prostate,

abdominal ailments treatment, and stomach ulcers treatment⁵. It also showed significant alpha amylase⁶ and acetyl cholinesterase (AChE) enzyme inhibitory activity⁷, the plant extract of *Paronychia argentea* showed antioxidant activity⁸. Other *in vivo* and *in vitro* studies on different extracts from *Paronychia argentea* revealed the immunomodulating activity of the plant⁹.



Many chemical compounds have been identified and isolated from *Paronychia argentea* as: jaceosidin, tricrin, nepetin, octadecanoic acid, 1-docosanol, glycerol octadecanoate monoester, β -sitosteryl glucoside, β -sitosterol, and luteolin-3- methyl ether, ethyl- α -D-galacto-pyranoside, D-pinitol(Pinitol is cyclitol)and vanillic acid, luteolin¹⁰. Vanillic acid and luteolin are considered as the major's active constituent of *Paronychia argentea* responsible for its biological activity¹¹.



MATERIAL AND METHODS

Materials

Plant material of *P. argentea*, aerial part, was collected on April 2016 from Ajloun area, Jordan. The dried plant material (600g) was grounded and soaked in ethanol (90%) for three weeks with frequent agitation, the alcohol solution was and then filtered and evaporated using Rota Vapor to obtain a solid residue (48 g).

Standard luteolin Primary Reference Standard, batch number HWI01784 was purchased from HWI Pharma services GmbH–Germany. Standard vanillic acid Primary Reference Standard, lot number STBD6012V was purchased from Sigma Aldrich – Germany, Distilled water, Acetonitrile HPLC grade, Methanol HPLC grade, Acetic Acid 99% HPLC grade.

METHODS

High Pressure Liquid Chromatographic analysis Preparation of Mobile phase

Distilled water, methanol and acetic acid were mixed at a ratio of (700:300:10) ml respectively then passed through a nylon membrane filter having a pore size of 0.45 μ m and sonicate to degas.

Preparation of Reference Standards

3.0 mg of Luteolin RS were accurately weighed and transferred to a 5 ml volumetric flask

then 3 ml of mobile phase were added and sonicate for 10 minutes. Mobile phase was added to complete the volume to 5ml (Final Concentration 0.60 mg/ml). In a similar way vanillic acid RS solution using was prepared.

Accurately weighed a 20 mg of Vanillic acid RS and transferred to a 50-ml volumetric flask then 30 ml of mobile phase was added, and the solution was sonicated for 10 minutes and made up to volume with the mobile phase(Final Concentration 0.4 mg/ml).

Chromatographic condition

All analyses were performed using an HPLC HITACHI Chromaster (5160 HPLC pump) & (5410, dual λ UV absorbance detector), HPLC column ODS-3 (150X4.6) mm, (5 μ m) and 5310 column oven and auto sampler 5260 were used during the HPLC analysis.

Standard solutions were injected using equal volumes of 20 μ l of at a flow rate of 1.0ml/min. and a wavelength of 260 nm.

To calculate the percentage of both vanillic acid and luteolin in the sample taken of the dry extract, the following formula was used.

$$\frac{AUC_{Sample}}{AUC_{STD}} \times \frac{C_{STD} \times P}{C_{Sample}} \times 100$$

Where:

- AUC_{sample} : area of sample.
 AUC_{STD} : area of standard.
 C_{STD} : Concentration standard taken (μ g/ml).
 P : Potency of Luteolin standard (91.82%)
 P : Potency of Vanillic acid (97%)
 C_{sample} : Concentration of Sample taken (μ g/ml).

Sample preparation for HPLC analysis

4.0 g of dry extract were transferred to a 25 ml volumetric flask, and 15 ml diluent (mobile phase) was added and sonicated for 20 minutes. The volume was completed with diluent (mobile phase) and filtered through 0.45 μ m Nylon filter.

Method Validation

The analytical method used for the quantification of Luteolin and Vanillic acid in the

P. argentea extracts was validated for linearity, LOD and LOQ, precision, as previously described.

RESULTS AND DISCUSSION

Optimization of Chromatographic Conditions

The HPLC conditions were optimized for the mobile phase composition, column temperature, wavelength, and flow rate (Table below). Detection wavelengths were set according to the ultraviolet (UV) absorption maxima of the compounds (260 nm).

Chromatographic conditions	
Column	ODS-3 (150X4.6) mm, 5 μ m
Column Oven	30°C
Flow rate	1.0 ML/min.
Injection volume	20 μ l
Wavelength	260 nm
Flushing solution	Methanol and distilled water at a ratio of 1:1
Runtime	15 min.

Peak of Luteolin and Vanillic acid standard preparations appear at retention time of 1.3 min 5.7 min. respectively as shown in Fig. 5 and Fig. 6 respectively, and a sample preparation was injected into the chromatograph and two peaks appear at retention times of 1.6 min. 5.7 min. respectively as shown in Figure 7.

Determination of major components in *Paronychia argentea* Lam dry extract

It was found that the Luteolin content is 0.4% while vanillic acid content is 0.1%

Table 1: Standard curve of assay vanillic acid standard. (Nominal concentration= 0.4 mg/ml)

Injection(μ L)	Concentration (μ g/ml)	Area	Average	RSD
100	199.0000	39927850	39775006	0.54
85	174.1250	39622161	34480770	0.10
75	149.2500	34530200	29111735	0.96
60	124.3750	29509963	24543949	0.41
50	99.5000	24400481	19482803	0.09
40	74.6250	19508796	14549070	0.11
25	49.7500	14525657	9651494	0.16
		96722831		

$R^2 = 0.99981$

Slope= 201133.9864

Intercept = -479056.6964

Method Validation

Linearity

The linearity of the assay method of vanillic acid was determined in the range from 25, 40, 50,60,75, 85 and100%, proportional to the concentration relative to the prescribed standard concentration 0.4 mg/ml and the curve was linear over a this large number of concentration and exhibited a linear regression ($R^2 = 0.99981$) and slope of 201133.9864.

Statistical analysis are shown in Table1 and a plot of area under the curve versus concentration can be seen in Fig. 1. The standard curve was plotted and evaluated for linearity.

The obtained equation for the standard curves was: Equation: $Y = AX - B$, Where B is the intercept with Y-axis and A is the slope

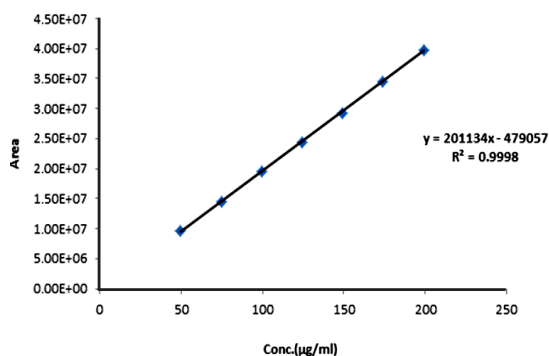


Fig. 1. Regression for concentrations vs. area of assay vanillic acid standard

Table 2: Standard curve of assay of Luteolin. (Nominal concentration= 0.6 mg/ml)

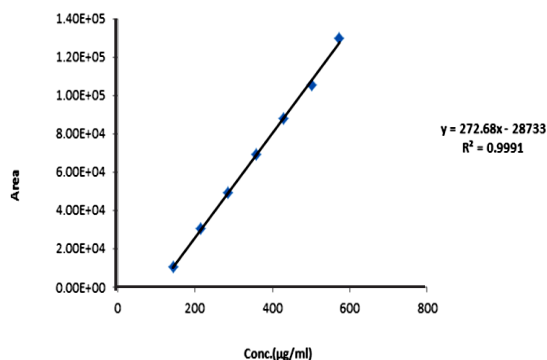
Injection(μ L)	Concentration(μ g/ml)	Area	Average	RSD
20	572.9568	129915 129030	129473	0.48
17.5	501.3372	105752 105580	105666	0.12
15	429.7176	88067 87818	87943	0.20
12.5	358.0980	69181 69262	69222	0.08
10	286.4784	49206 49853	49530	0.92
7.5	214.8588	30157 30514	30336	0.83
5	143.2392	10214 10234	10224	0.14

 $R^2 = 0.99908$

Intercept=

Slope= 272.6805

-28733.2679

**Fig. 2. Regression for concentrations vs. area of assay Luteolin standard****Table 3: System precision for vanillic acid standard**

Area under the curve for 10 replicate injections	
Injection #	Standard area of Vanillic Acid
1.	110246472
2.	111954152
3.	109265151
4.	110448955
5.	112542944
6.	111111762
7.	110796209
8.	108973254
9.	111229759
10.	110073105
Average	110664176.3
RSD	0.98

Precision**System repeatability**

In this test, system repeatability tests were examined and standard solutions containing Vanillic acid were Prepared, and injected 10 times into the HPLC system. Average of peak areas and % RSD values were calculated and shows to be within acceptable limits of 0.98%. The resulting areas and RSD values shown in Table 3.

Analysis Repeatability

For the determination of the repeatability of the extract, six samples obtained from multiple sampling of extract were analyzed, in a single laboratory on a single day. Assay percentage, and RSD values were calculated. The results obtained are listed in Table 4.

Table 4: Analysis repeatability extract

	Sample #	Assay% of Vanillic Acid	Assay % of Luteolin
Dry Extract	1.	0.13	0.43
	2.	0.13	0.42
	3.	0.14	0.47
	4.	0.13	0.42
	5.	0.14	0.46
	6.	0.13	0.41
	Average	0.13	0.44
RSD %	3.5	5.1	

Range

Range is the concentration interval between

the upper and lower concentration of analyte for which is shown that the method has suitable level of linearity. Range for determination is extended from 25% to 100% vanillic acid.

Limit of Detection

Limit of detection (LOD) is the lowest concentration of analyte in a sample, which can be detected, but not necessary quantified, under the stated experiment conditions.

For vanillic acid it is estimated by extrapolation of regression line through Y axis from standard curve at low concentration as in the following expression.

The LOD was found to be 0.40 ($\mu\text{g/ml}$) for vanillic acid which is equivalent to 0.2% and 1.2($\mu\text{g/ml}$) which is equivalent to 0.6% of nominal value of Luteolin respectively.

Limit of Quantitation

The limit of quantitation (LOQ) is the lowest concentration of analyte in a sample, which can be determined with an acceptable accuracy and precision detected.

For Vanillic acid is estimated by extrapolation of regression line through Y axis from standard curve at low concentration as in the following expression Figure 3 and 4.

Table 5: Standard solution of vanillic acid prepared for calculation of LOD and LOQ

Injection(μL)	Concentration ($\mu\text{g/ml}$)	Area	Average	RSD
10	99.5000	19482803 19508796	19495800	0.09
7.5	74.6250	14549070 14525657	14537364	0.11
5.0	49.7500	9651494 9672831	9662163	0.16

$R^2 = 0.99998$ Slope = 197661.0452
 Intercept = -185347.0000 SD of Intercept = 24027.8748

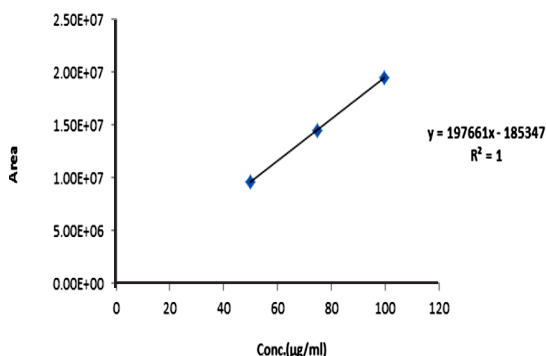


Fig. 3. Regression for concentrations vs. area of vanillic acid standard for LOD and LOQ

Table 6: LOD and LOQ of vanillic acid estimation results

Parameter	Concentration	Percentage from Nominal
LOD	0.40 ($\mu\text{g/ml}$)	0.2%
LOQ	1.2 ($\mu\text{g/ml}$)	0.6%

The LOQ was found to be 1.2 ($\mu\text{g/ml}$) for vanillic acid which is equivalent to 0.6% of nominal value and 9.7($\mu\text{g/ml}$) which is equivalent to 4.8% of nominal

value of Luteolin respectively (Tables 6, 7 and 8).

Filter compatibility

Variation of the filter type is described below and the results are reported in Table 9.

Several attempts were used to develop a reverse phase HPLC method of analysis for the separation of the two main components luteolin and vanillic acid available in dry extract of Paronychia.

A suitable mobile phase was established to separate the two major compounds of the extract in one run with a suitable and reasonable time, where both vanillic acid and luteolin were detected at a wavelength of 260 nm, and a sample preparation was injected into the chromatograph and two peaks appear at retention times of 1.3 min. 5.7 min. respectively and a runtime of 15 min. and at a flow rate of 1.0 ml/min. using ODS-3 (150X4.6) mm, 5 μm .

A baseline resolution was obtained under the testing of the analysis conditions, as well as

the chromatograms of the standards and samples preparation are shown in Fig. 5, 6 and 7. this method has been validated as per ICH guidelines for linearity, precision, limit of detection and limit of quantitation for both main active components, and a linear relationship was noticed for the two compounds and the correlation coefficient was $R^2 = 0.9998$. method developed and used is precise, repeatability LOD, LOQ and filter compatibility, it can be concluded that

the method developed and used is precise, sensitive accurate and reproducible, and facilitate quantitative determination of this extract and will accomplish the objective of this study and consequently utilizing a local Jordanian plant since the active materials in the extract are variable. The concentration of luteolin and vanillic acid in the dry extract was 0.4 and 0.1% respectively.

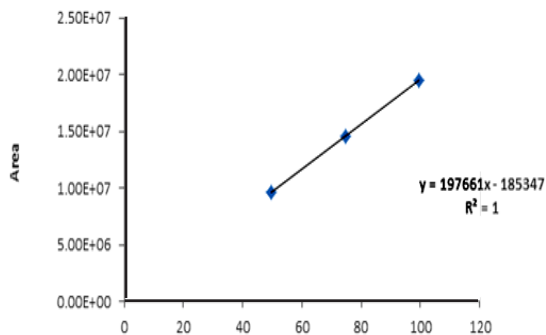


Fig. 4. Regression for concentrations vs. area of assay vanillic acid Luteolin standard

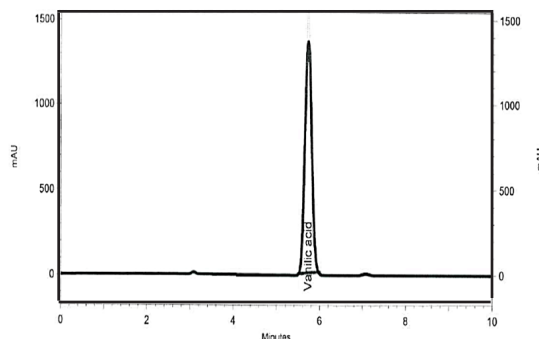


Fig. 6. Represent the chromatogram of vanillic acid

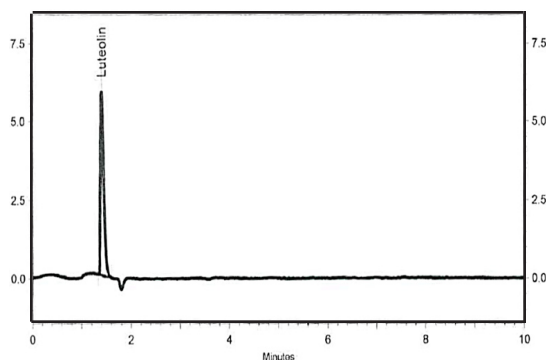


Fig. 5. Represent the chromatogram of Luteolin Reference Standard

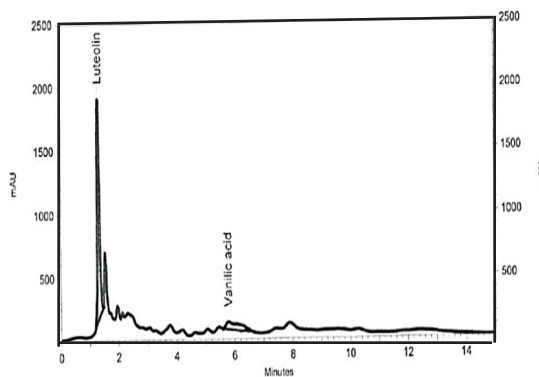


Fig.7. Represent the chromatogram of Luteolin and Vanillic Acid in dry extract sample

Table7: Standard solution of luteolin prepared for calculation of LOD and LOQ

Linearity of Luteolin for LOD & LOQ				
Injection (µL)	Concentration (µg/ml)	Area	Average	RSD
10	286.4784	49206	49530	0.92
		49853		
7.5	214.8588	30157	30336	0.83
		30514		
5	143.2392	10214	10224	0.14
		10234		
R ² =0.99982		Slope= 274.4046		
Intercept =-28928.5833		SD of intercept = 264.8594		
LOD Conc.= 3.19		LOD %= 1.6		
LOQ Conc.=9.7		LOQ%=4.8		

Table 8: LOD and LOQ of Luteolin estimation results

Parameter	Concentration	Percentage from Nominal
LOD	3.19 (µg/ml)	1.6%
LOQ	9.7 (µg/ml)	4.8%

Table 9: Variation of the filter type is described filter compatibility results

Vanillic acid		
Filter type	Area of Standard	Change % from unfiltered
Unfiltered	110448955	-
0.45µm Glass	112401742	101.8
0.45 µm Nylon	111021145	100.5
0.45µm PTFE	112251423	101.6

Table 10: R_f values, areas under the curve and percentage of the area for Luteolin and Vanillic acid

Peak Number	Ingredient name	R _f	Area	Theoretical plates	Asymmetry	Area%
1	Luteolin	1.3	33798868	1903	1.2	79.0
2	Vanillic acid	5.7	8971344	511	3.1	21.0

CONCLUSION

The two major peaks of Luteolin and Vanillic acid were simultaneously separated and determined successfully from the other components in the *Paronychia argentea* Lam dry extract collected from the plant located in Jordan and was found that the Luteolin content is 0.4% while vanillic acid content is 0.1% using a validated chromatographic method

of analysis. These two compounds are considered as a standardized reference of the plant material.

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REFERENCES

1. AlRawashdeh, I. and AlRawashdeh, N. *Biology Agriculture healthcare.*, **2016**, 6, 128.
2. Al-Khalil, S. *International Journal of Pharmacognosy*, **1995**, 33, 317.
3. Ahmed, S.; Mallick I. and Hasan, M. *J. Pharmacogn. Phytochem.*, **2017**, 6, 1780-1787.
4. Afifi, F.; Al-Khalidi, B. and Khalil, E. *J Ethnopharmacol.*, **2005**, 100(3), 314-8.
5. Hamdan, I. and Afifi F. *J Ethnopharmacol.*, **2004**, 93(1), 117-21.
6. Ali-Shtayeh M.; Yaniv, Z. and Mahajna J. *J Ethnopharmacol.*, **2000**, 73(1-2), 221-32.
7. Ferreira, A.; Proença, C.; Serralheiro, M.; Araújo, M. *J Ethnopharmacol.*, **2006**, 108(1), 31-7.
8. Tawaha, K.; Alali, F. Q.; Gharaibeh, M.; Mohammad, M.; El-Elimat, T. *Food Chemistry.*, **2007**, 104, 1372.
9. Szallasi, A. *Drug Aging.*, **2001**, 18, 561.
10. Baqi, Y. N. H., University of Jordan. MSc Thesis., **2003**.
11. Vriens, J.; Appendino, G.; Nilius, B. *Molecular Pharmacology.*, **2009**, 75, 1262.